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Key indicators

Single-crystal X-ray study T = 183 KMean $\sigma(C-C) = 0.005 \text{ Å}$ R factor = 0.068 wR factor = 0.097 Data-to-parameter ratio = 12.7

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

Dimethyl{3-[1-methyl-4-(1-methyl-4-nitropyrrole-2-carboxamido)pyrrole-2-carboxamido]propyl}ammonium chloride methanol solvate

In the title compound, $C_{17}H_{25}N_6O_4^+ \cdot Cl^- \cdot CH_4O$, the cation adopts a bowed conformation with π -conjugation *via* two pyrrole rings and and two peptide groups.

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Comment

All living organisms (excluding certain viruses) store their genetic information in the form of double-helical DNA. Biochemical access to this information is based on specific protein-DNA interactions. Despite recent progress using biological selection methods (phage display), predictive chemical principles for protein-DNA recognition are still considered complex (Greisman & Pabo, 1997; Choo & Klug, 1997). Thus, this problem has prompted chemists to synthesize small molecules relevant to the problem of DNA recognition. Inspired by the natural products netropsin and ditamycin, which bind to the AATT and AAATT sequences, respectively, in the minor groove of DNA, chemists have recently developed polyamides containing N-methylpyrrole and N-methylimidazole amino acids which can recognize and bind in the minor groove of predetermined DNA sequences with high affinity and specificity comparable to naturally occurring DNA-binding proteins (Trauger et al., 1996; Swalley et al., 1997; Turner et al., 1997; Trauger et al., 1998; Dervan & Buril, 1999). The co-crystal structures of netropsin, ditamycin and some polyamides showed that the polyamides exerted an influence on DNA structures, and DNA also influenced the structure of the polyamides (Berman et al., 1979; Kopka et al., 1985; Collect et al., 1987; Balendiran et al., 1995; Kielkopf, Baird et al., 1998; Kielkopf, White et al., 1998; Shi et al., 2002). To understand the principle of the interaction, which is significant for the design of new DNA-binding compounds, more structural information is needed on polyamide-DNA and polyamides themselves. In this paper, we discuss the structure of the title polyamide, which contains two Nmethylpyrrole rings.



© 2003 International Union of Crystallography Printed in Great Britain – all rights reserved The overall appearance of the cation in the title compound, (I), is a bowed molecule (Fig. 1). The bond lengths and angles





The structure of the cation in the title compound, (I), with displacement ellipsoids drawn at the 50% probability level for non-H atoms.





The packing diagram of (I), viewed along the a axis. Hydrogen bonds are shown as dashed lines.

are not unusual (Table 1). There is conjugation involving the two pyrrole rings and the two peptide groups. Their atoms are essentially coplanar, forming a plane consisting of 22 atoms (N2–N6, O1–O4 and C3–C15). The maximum deviations from the plane are 0.250 (2) and -0.247 (2) Å for O1 and O2, and the minimum deviations only 0.001 (3) and -0.003 (3) Å for C13 and N5, respectively. The dihedral angle between the two pyrrole rings is $3.7 (2)^{\circ}$. An obvious difference from a similar compound, such as netropsin, is a smaller rotation [8.7 (5)°] about the C10–C11 bond than that of netropsin (26° rotation about the C6–C9 bond); this causes the two pyrrole groups of netropsin to be skewed with respect to one another (Berman *et al.*, 1979). The dihedral angle between the two peptide units is 16.9 (2)°. This is larger than that between the

two pyrrole rings, and may be due to one peptide bond being linked to an unconjugated dimethylaminopropyl group.

Hydrogen-bond parameters are listed in Table 2 and a packing diagram is shown in Fig. 2. In the crystal packing, the molecules are held together by a number of intermolecular hydrogen bonds involving atoms N1, N2 and N4 of the cation, as well as the O atom of the solvent methanol molecule and the chloride anion.

Experimental

The literature procedure, slightly modified, for the preparation of 3-[1-methyl-4-(1-methyl-4-nitropyrrole-2-carboxamido)pyrrole-2-carboxamido]dimethylaminopropane was followed (Nishiwaki *et al.*, 1988). The resulting compound was dissolved in methanol (100%) with a few drops of 1 N HCl. The solution was left at 277 K and crystals of (I) appeared after several months.

Crystal data

$C_{17}H_{25}N_6O_4^+ \cdot Cl^- \cdot CH_4O$	$D_x = 1.374 \text{ Mg m}^{-3}$
$M_r = 444.92$	Mo $K\alpha$ radiation
Monoclinic, $P2_1/c$	Cell parameters from 1027
a = 6.2129 (13) Å	reflections
b = 17.801 (4) Å	$\theta = 2.4 - 19.6^{\circ}$
c = 19.442 (4) Å	$\mu = 0.22 \text{ mm}^{-1}$
$\beta = 90.765 \ (4)^{\circ}$	T = 183 (2) K
V = 2150.1 (8) Å ³	Block, yellow
Z = 4	$0.20 \times 0.20 \times 0.20$ mm
Data collection	
SMART 1K CCD area-detector	2335 reflections with $I > 2\sigma(I)$

 $R_{int} = 0.074$ $\theta_{max} = 24.8^{\circ}$ $h = -7 \rightarrow 6$ $k = -20 \rightarrow 19$

 $-22 \rightarrow 21$

diffractometer
ω scans
Absorption correction: none
8201 measured reflections
3529 independent reflections

Refinement

Refinement on F^2	H-atom parameters constrained
$R[F^2 > 2\sigma(F^2)] = 0.068$	$w = 1/[\sigma^2(F_o^2) + (0.0108P)^2]$
$vR(F^2) = 0.097$	where $P = (F_o^2 + 2F_c^2)/3$
S = 0.98	$(\Delta/\sigma)_{\rm max} < 0.001$
3529 reflections	$\Delta \rho_{\rm max} = 0.19 \ {\rm e} \ {\rm \AA}^{-3}$
278 parameters	$\Delta \rho_{\rm min} = -0.21 \text{ e } \text{\AA}^{-3}$

Table 1

Selected geometric parameters (Å, °).

C1-N1	1.499 (4)	C10-O2	1.227 (4)
C1-C2	1.514 (5)	C10-N4	1.337 (4)
C2-C3	1.499 (4)	C10-C11	1.490 (4)
C3-N2	1.448 (4)	C11-C12	1.359 (5)
C4-O1	1.225 (4)	C11-N5	1.387 (4)
C4-N2	1.351 (4)	C12-C13	1.390 (4)
C4-C5	1.470 (5)	C13-C14	1.372 (5)
C5-C6	1.368 (4)	C13-N6	1.430 (4)
C5-N3	1.381 (4)	C14-N5	1.355 (4)
C6-C7	1.389 (5)	C15-N5	1.463 (4)
C7-C8	1.387 (5)	C16-N1	1.489 (4)
C7-N4	1.411 (4)	C17-N1	1.488 (4)
C8-N3	1.368 (4)	N6-O4	1.225 (3)
C9-N3	1.460 (4)	N6-O3	1.247 (4)
N2-C4-C5-C6	12.2 (5)	C5-C4-N2-C3	-178.6(3)
N4-C10-C11-C12	-8.7(5)	O2-C10-N4-C7	-0.7(5)
O1-C4-N2-C3	2.0 (5)	C11-C10-N4-C7	179.2 (3)

Table 2	
Hydrogen-bonding geometry (Å, °).	

$D - H \cdot \cdot \cdot A$	D-H	$H \cdot \cdot \cdot A$	$D \cdots A$	$D - \mathbf{H} \cdot \cdot \cdot A$
$O5-H5\cdots Cl1^i$	0.84	2.34	3.152 (3)	162
$N1-H1\cdots Cl1^{ii}$	0.85	2.28	3.099 (3)	161
N4-H4···O5 ⁱⁱⁱ	0.86	2.22	3.060 (4)	164
$N2-H2\cdots Cl1^{iv}$	0.77	2.72	3.430 (3)	154

Symmetry codes: (i) 1 + x, y, z; (ii) -x, 1 - y, 1 - z; (iii) 2 - x, 1 - y, 1 - z; (iv) 1 - x, 1 - y, 1 - z.

H atoms attached to C and O atoms were placed in geometrically idealized positions, with $Csp^2-H = 0.95$ Å, $Csp^3-H = 0.98$ Å, $Csp^3-H = 0.98$ Å, and $Osp^3-H = 0.84$ Å, and constrained to ride on their parent atoms, with $U_{\rm iso}(H) = 1.2U_{\rm eq}(C)$ and $U_{\rm iso}(H) = 1.5U_{\rm eq}(O)$. H atoms on N atoms were located in a difference Fourier map and refined with a common $U_{\rm iso}$ value. The N-H distances are in the range 0.77–0.86 Å.

Data collection: *SMART* (Bruker, 2000); cell refinement: *SAINT* (Bruker, 2000); data reduction: *SAINT*; program(s) used to solve structure: *SHELXS*97 (Sheldrick, 2000); program(s) used to refine structure: *SHELXL*97 (Sheldrick, 2000); molecular graphics: *SHELXTL/PC* (Sheldrick, 1999); software used to prepare material for publication: *SHELXTL/PC*.

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References

- Balendiran, K., Rao, S. T., Sekharudu, C. Y., Zon, G. & Sundaralingam, M. (1995). Acta Cryst. D51, 190–198.
- Berman, H. M., Neidle, S., Zimmer, C. & Thrum, H. (1979). Biochem. Biophys. Acta, 561, 124–131.
- Choo, Y. & Klug, A. (1997). Curr. Opin. Struct. Biol. 7, 117-125.
- Bruker (2000). *SMART* (Version 5.0) and *SAINT* (Version 6.02). Bruker AXS Inc., Madison, Wisconsin, USA.
- Collect, M., Frederick, C. A., Wang, A. H.-J. & Rich, A. (1987). Proc. Natl Acad. Sci. USA, 84, 8385–8389.
- Dervan, P. B. & Buril, R. W. (1999). Curr. Opin. Chem. Biol. 3, 688-693.
- Greisman, H. A. & Pabo, C. O. (1997). Science, 275, 657-661.
- Kielkopf, C. L., Baird, E. E., Dervan, P. B. & Rees, D. C. (1998). Nat. Struct. Biol. 5, 104–109.
- Kielkopf, C. L., White, S., Szewczyk, J. W., Turner, J. M., Baird, E. E., Dervan, P. B. & Rees, D. C. (1998). *Science*, **282**, 111–115.
- Kopka, M. L., Yoon, C., Goodsell, D., Pjura, P. & Dickerson, R. E. (1985). Proc. Natl Acad. Sci. USA, 82, 1376–1380.
- Nishiwaki, E., Tanaka, S., Lee, H. & Shibuya, M. (1988). *Heterocycles*, 27, 1945–1952.
- Sheldrick, G. M. (1999). *SHELXTL/PC*. Version 6.10. Bruker AXS Inc., Madison, Wisconsin, USA.
- Sheldrick, G. M. (2000). *SHELXS*97 and *SHELXL*97. University of Göttingen, Germany.
- Shi, K., Mitra, S. N. & Sundaralingam, M. (2002) Acta Cryst. D58, 601-606.
- Swalley, S. E., Baird, E. E. & Dervan, P. B. (1997). J. Am. Chem. Soc. 119, 6953–6961.
- Trauger, J. W., Baird, E. E. & Dervan, P. B. (1996). *Nature (London)*, **382**, 559–561.
- Trauger, J. W., Baird, E. E. & Dervan, P. B. (1998). Angew. Chem. Int. Ed. 37, 1421–1423.
- Turner, J. M., Baird, E. E. & Dervan, P. B. (1997). J. Am. Chem. Soc. 119, 7636–7644.